ORIGINAL ARTICLE

Antibiotic Susceptibility Pattern of Bacterial Isolates from Wound Infections in a Tertiary Care Hospital

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ABSTRACT

Introduction: The human skin and soft tissue infections (SSTIs) caused by microbial pathogens during or after trauma, burn injuries, and surgical procedures result in the production of pus. Wound infections which commonly occur under hospital environment result in significant morbidity, prolonged hospitalization, and huge economic burden. The emergence of antibiotic resistance and its rapid spread among bacterial isolates are considered as grave threats to the public health worldwide. The objective of this study was to characterize the pyogenic bacteria from pus samples and to determine their antibiotic susceptibility

Materials and Methods: Various samples received from inpatients of different surgical wards over a period of one year from January 2016 to December 2016 were included in study, Samples were processed for Gram staining and culture. The samples were aseptically inoculated on blood agar and MacConkey agar plates, incubated aerobically at 35°C–37°C for 24–48 h. Identification and characterization of isolates were performed using standard microbiological methods. Antibiotic susceptibilities of bacterial isolates were determined according to the method recommended by the Clinical and Laboratory Standards Institute.

Results: Of the 2045 samples received, a total of 1028 isolates were obtained. Culture positivity of pus samples was 50.2%. *E. coli* (33.8%) was the most frequent pathogen followed by *K. pneumoniae* (20.4%), *S. aureus* (19.2%), *P. aeruginosa*(9.3%). Gram-negative isolates were predominant isolates as compared to Gram-positive isolates. Antibiogram results showed that *E. coli* and Klebsiella spp. were more resistant to cephalosporins, ciprofloxacin while being least resistant to imipenem, tigecycline. *P. aeruginosa* was more susceptible to piperacillin tazobactam, amikacin, imipenem. All isolates of *S. aureus* were susceptible to vancomycin (100%) & linezolid (100%).

Conclusion *E. coli* was the commonest isolate followed by *K. pneumoniae, S. aureus, P. aeruginosa,* Bacterial isolates exhibited high to moderate levels of resistance against different classes of antibiotics. The susceptibility data may be helpful in implementing empiric treatment strategies for pyogenic infections.

KEYWORDS: Wound infection, Gram positive, Gram negative, Susceptibility

INTRODUCTION

Skin abrasions due to surgical procedures, trauma, burns, diseases, and other factors affect this first line of defense and leads to microbial contamination resulting in infections.¹ The human skin and soft tissue infections (SSTIs) caused by microbial pathogens during or after trauma, burn injuries, and surgical procedures results in the production of pus, a white to yellow fluid produced during an inflammatory pyogenic infection due to bacteria which is comprised of dead WBCs, cellular debris, and necrotic

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The Gulf Journal of Dermatology and Venereology

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In spite of advances in control of infections, wound infections have not been completely controlled due to many reasons. The overall incidence of wound sepsis in India is from 10% to $33\%.^{5,6}$

Both aerobic and anaerobic bacteria have been implicated in wound infections which commonly occur under hospital environment and result in significant morbidity, prolonged hospitalization, and huge economic burden.⁷

Knowledge of these causative agents of wound infection has proven to be helpful in the selection of appropriate antimicrobial therapy and on infection control measures taken in health institutions.⁸

Fairly consistent studies have always been done all over the world to show a predictable bacterial profile and the antibiogram in their respective areas. This makes an important observation for a clinician who intends to give empirical treatment to his patients while laboratory culture reports are awaited.⁹

But despite advances in methods to control infections, wound infections have not been completely prevented due to the problem of drug resistance.¹⁰ The widespread uses of antibiotics, together with the length of time over which they have been available, have led to major problems of resistant organisms contributing to morbidity and mortality.¹¹ But on account of erratic use, malpractices or for natural causes, in recent years, drug resistance to many human pathogenic bacteria is being commonly reported from all over the world.¹²

During the last few decades, multidrug-resistant Gram-negative bacterial strains such as *Acinetobacter baumannii*, *E. coli*, *Klebsiella pneu-* *moniae, Pseudomonas aeruginosa,* and Grampositive methicillin-resistant *Staphylococcus aureus* (MRSA) were increasingly associated with pus infections under hospital settings due to extensive misprescription and inadequate dose regimen of antibiotics.¹³⁻¹⁵ Rapid emergence of multidrug-resistant bacteria poses a serious threat to public health globally due to the limited treatment options and lukewarm discovery of new classes of antibiotics.^{15,16}

The objective of this study is to characterize the pyogenic bacteria from wound samples and to determine their antibiotic susceptibilities to various antibiotics commonly used.

MATERIAL AND METHODS

Pus, necrotic tissue and swabs from wound infections received from inpatients of different surgical wards over a period of one year from January 2016 to December 2016 were included in study. The samples were processed for Gram staining and culture. The samples were aseptically inoculated on blood agar and MacConkey agar plates, incubated aerobically at 35°C–37°C for 24–48 h. Identification and characterization of isolates were performed using Gram staining, motility and various biochemical reactions.¹⁷

Antibiotic susceptibility of bacterial isolates was determined according to the method recommended by the Clinical and Laboratory Standards Institute. Inoculum (0.5 McFarland) was prepared and lawn culture was done on Muller-Hinton agar plates. Antibiotic discs were placed and plates were incubated at 37°C for 24 h. The zones of inhibition were measured. Isolates were classified as sensitive, intermediate, and resistant according to CLSI guidelines.(18). Isolates were characterized into MDR and XDR.

RESULTS

Of the 2045 pus samples received, a total of 1028 isolates were obtained. Culture positivity of pus samples was 50.2%. Gram-negative isolates (77.2%) were predominant isolates from pus samples as compared to Gram-positive isolates (22.8%). E. coli (33.8%) was the most frequent pathogen followed by K. pneumoniae (20.4%), S. aureus (19.2%), P. aeruginosa (9.3%). (Fig. 1) Antibiogram results showed that E. coli and Klebsiella isolates were more resistant to cephalosporins, ciprofloxacin while being least resistant to imipenem, tigecycline. P. aeruginosa was more susceptible to piperacillin tazobactam, amikacin, imipenem. (Fig. 2) Majority of Gram negative isolates were MDR (83.5%), 25.3% isolates were XDR. All isolates of S. aureus were susceptible to vancomycin (100%) and linezolid (100%), followed by 87.2% susceptibility to gentamicin, >70%

to erythromycin & clindamycin, 55.6% to ciprofloxacin, 50% to amikacin & cotrimoxazole and 21.4% to penicillin. Out of 196 *S. aureus* isolates, 52 (26.5%) were MRSA (Table 1)

DISCUSSION

In our study, a total of 1028 isolates were obtained from 2045 samples received. Gramnegative isolates were predominant isolates from pus samples as compared to Gram-positive isolates. *E. coli* (33.7%) was the most frequent pathogen followed by *K.pneumoniae* (20.4%), *S. aureus* (19.1%), *P. aeruginosa*(9.3%).

A study from Andhra Pradesh reported 40% Gram-positive cocci and 60% Gram-negative isolates in their study. *E. coli* followed by Klebsiella was also the most common Gramnegative bacteria isolated from the pus samples in their study.¹⁹

Such Gram-negative bacterial dominance in the

Gram Negative org. (n=793)	Total	%	AMP	AMK	GEN	сот	СРМ	CTX/ CAZ	CIP	TZP	IPM	ERT	MEM	AZT	TGC	COL
E.coli	374	33.7	-	86.4	62.5	60.2	33.7	18.7	21.3	63.4	85.8	48.9	68	-	98.5	100
Klebsiella ssp.	210	20.4	-	50	39.5	44.7	26.19	13.8	23.3	44.7	74.2	31.4	60	-	90.4	62.3
Pseudomonas spp.	96	9.3	-	79.1	60.4	-	76	60.4	57.2	77	75	-	64.5	52	-	65.6
Acinetobacter	75	7.3	-	13.3	10.6	10.6	9.3	8	8	21.3	36	9.3	21.3	-	94.6	65.3
Others	65	6.3														
Gram positive org. (n=793)	Total	%	PEN	AMK	GEN	сот	СІР	ТЕТ	STR	AMC	ERY	CLI	FOX	TEC	VAN	LZ
S. aureus	196	19.1	21.4	50	87.2	50	55.6	-	-	-	74.4	75.5	73.4	100	95.4	98.4
Enterococcus	39	3.7														

Table 1 Antibiotic Susceptibility profile of bacterial isolates(n=1028)Antibiotic susceptibility profile of isolates from wound infections

(Ampicillin-AMP, Amikacin-AMK, Gentamicin-GEN, Cotrimoxazole-COT, Ceftazidime-CAZ, Cefepime-CPM, LEVO-Levofloxaxin, MIN-Minocycline. Ciprofloxacin-CIP, Piperacillin/tazobactum-TZP, Cefoperazone/Sulbactam-CSL, AMC-Amoxiclav, Imipenem-IPM, Meropenem-MEM, Ertapenem-ERT, DOR=Doripenem, Aztreonam-AO, Tigecycline-TGC, Penicillin-PEN, Daptomycin-DPT, GHL-Gentamycine (High), Tetracycline(TET) aerobic growth in pus culture has been highly seconded by studies reported by Ghosh et al.[18] and Zubair et al.²¹ Another study by Basu et al.²² also reported Pseudomonas and *E. coli* spp. to be the most commonly occurring pathogens in wound infections, in that order. Raza et al.²³ found *E. coli* to be the most common pathogen with similar observations by studies conducted in Nigeria.²⁴

Antibiogram results of Gram negative isolates from the present study showed that *E. coli* and Klebsiella isolates were more resistant to cephalosporins and ciprofloxacin while being least resistant to imipenem, tigecycline. *P. aeruginosa* was more susceptible to amikacin, piperacillin- tazobactam and imipenem.

A similar study from Andhra Pradesh revealed high resistance by Gram-negative bacteria to even fourth-generation cephalosporins. Whereas, carbapenems were still sensitive though increasing resistance was observed to meropenem. Resistance was seen by most of the isolates to quinolones. Combination drugs such as piperacillin+tazobactam and cefoperazone+sulbactum showed good amount of sensitivity.¹⁹ Similar studies by Javeed et al.,²⁵ Rao et al.,⁹ and Anguzu and Olila²¹ corroborated our findings.

S. aureus isolates in our study were most susceptible to vancomycin (100%) and linezolid (100%) and least susceptible to penicillin (21.4%). Similar high susceptibility was shown by *S. aureus* to high-end drugs such as linezolid and vancomycin in a study from Andhra Pradesh.¹⁹ These findings are similar to those of Rao et al.,⁹ who also found *S. aureus* to be resistant to penicillin (84.62%), erythromycin (84.62%), and sensitive to clindamycin (65.38%) and vancomycin (100%). Out of 196 *S. aureus* isolates, 26.5% were MRSA Unfortunately, this only shows that Staphylococcus has become highly resistant to the first and second lines of treatment.

CONCLUSIONS

Pyogenic wound infections were found prevalent and *E. coli* isolates were predominant followed by *K. pneumoniae*. Bacterial isolates exhibited high to moderate levels of resistance against different classes of antibiotics. The susceptibility data may be helpful in implementing empiric treatment strategies for pyogenic infections. Strict health policies should also be implemented to restrict the unsupervised antibiotic use as well as continuous monitoring and reporting of antibiotic resistance.

REFERENCES

- Howard RJ, Ravitch MM, Steichen FM. Host against Infections. Current Problems in Surgery. New Eng J Med. 1980; 12:1823-30.
- Cogen AL, Nizet V, Gallo RL. Skin microbiota: a source of disease or defence? Br J Dermatol. 2008; 158:442-55.
- Dryden MS. Complicated skin and soft tissue infection.Journal of Antimicrobial Chemotherapy. 2010; 65:iii35-44.
- Scalise A, Bianchi A, Tartaglione C, Bolletta E, Pierangeli M, Torresetti M, et al. Microenvironment and microbiology of skin wounds: the role of bacterial biofilms and related factors.Seminars in Vascular Surgery. 2015; 28:151-59.
- Akimoto Y, Mochizuki Y, Uma A, Omata H, Shibutani J, Nishimura H, et al. Amoxicillin concentration in pus from abscess caused by odontogenic infection. Gen Pharmacol 1994; 25:111-13.
- Koneman EW, Allen SD, Janda WM, Schreckenberger PC, Propcop GW, Woods GL, et al Philadelphia Color Atlas and Textbook of Diagnostic Microbiology,

6th edn. Philadelphia, PA: Lippincott Williams and Wilkins, 2005. pp. 624-62.

- BowlerPG, DuerdenBI, ArmstrongDG. Wound microbiology and associated approaches to wound management. Clinical Microbiology Reviews. 2001; 14:244-49.
- Adebayor OS, Deboye OK, Emiola AR. Wound Infections in two Health Institutions in IIE-Ife, Nigeria: Results of a cohort study. Osto WoundManag. 2003; 49:52-57.
- Rao DVMVSVR, Basu R, Biswas DB. Aerobic bacterial profile and antimicrobial susceptibility pattern of pus isolates in a south Indian tertiary care hospital. IOSR J Dent Med Sci. 2014; 13:59-62.
- Thomas KH. Surgical Wound Infection, an Overview. Am J Med. 1981; 70:712-18.
- Elmer WK, Stephen DA, William MJ, Schreckenberger PC, Winn WC. Antimicrobial Susceptibility testing in, colour atlas and textbook of Diagnostic Microbiology. 5th edition. Philadelphia: Raven Publisher; 1997:69-120.
- Jahan F, Lawrence R, Kumar V, Junaid M. Evaluation of antimicrobial activity of plant extracts on antibiotic susceptible and resistant Staphylococcus aureus strains. J Chem Pharma Res. 2011; 3:777-89.
- Rice LB. Antimicrobial resistance in gram-positive bacteria. The American Journal of Medicine. 2006; 119:S11-9.
- MisicAM, GardnerSE, GriceEA. The Wound Microbiome: modern approaches to examining the role of microorganisms in impaired chronic wound healing. AdvancesIn Wound Care. 2014; 3:502-10.
- 15. IredellJ, BrownJ, TaggK. Antibiotic resistance in Enterobacteriaceae: mechanisms and clinical implications.BritishMedical Journal.2016; 352:6420.
- 16. CerceoE, DeitelzweigSB, ShermanBM, AminAN. Multidrug-resistant gram-negative bacterial infections in the hospital setting: overview, implications for clinical practice, and emerging treatment options. Microbial Drug Resistance. 2016; 22:412-31.

- Collee JG, Marr W. Specimen collection, culture containers and media. In: Collee JG, Marmion BP, Fraser AG, Simmons A, eds. Mackie & McCartney Practical Medical Microbiology. 14th ed. Elsevier; 2014. p.95-112.
- CLSI. Performance standards for Antimicrobial Susceptibility testing. 27th ed. CLSI supplement M100. Wayne,PA: Clinical and Laboratory standards Institute, 2017.
- Mantravadi HB, Chinthaparthi MR, Shravani V. Aerobic isolates in pus and their antibiotic sensitivity pattern: a study conducted in a teaching hospital in Andhra Pradesh. Int J Med Sci Public Health. 2015;4:1076-79.
- Ghosh A, Karmakar PS, Pal J, Chakraborty N, Debnath NB, Mukherjee JD. Bacterial incidence and antibiotic sensitivity pattern in moderate and severe infections in hospitalized patients. J Indian Med Assoc. 2009; 107:21-2, 24-5.
- Zubair M, Malik A, Ahmad J. Clinico-microbiological study and antimicrobial drug resistance profile of diabetic foot infections in north India. Foot (Edinb). 2011; 21:6-14.
- Basu S, Ramchuran Panray T, Bali Singh T, Gulati AK, Shukla VK. A prospective, descriptive study to identify the microbiological profile of chronic wounds in outpatients. Ostomy Wound Manage. 2009; 55:14-20.
- Raza MS, Chander A, Ranabhat A. Antimicrobial susceptibility patterns of bacterial isolates in postoperative wound infections in a tertiary care hospital, Kathmandu, Nepal. Open J Med Microboil. 2013; 3:159-63.
- Taiwo SS, Okesina AB, Onile BA. In vitro antimicrobial susceptibility pattern of bacterial isolates from wound infections in University of Ilorin teaching hospital. Afr J Clin Exp Microbiol. 2002; 3:6-10.
- 25. Javeed I, Hafeez R, Anwar MS. Antibiotic susceptibility pattern of bacterial isolates from patients admitted to a tertiary care hospital in Lahore. Biomedica. 2011; 27:19-23.